

Perspectives on Homology and Similarity Searching

William Pearson
U. of Virginia

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Perspectives on Homology and Similarity Searching

- A brief (150+ years) of “Homology” as a concept
- The break-throughs: “Homology” (similarity searching) as a tool for discovery
- Why (and when) do we infer “Homology” from “similarity” (hint: *excess similarity*)
- What mistakes, and when?
- Trust your *positive* BLAST search results

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A bit about me - 1

- Undergraduate at the U. of Illinois Urbana, 1967-1971. Major in Chemistry, with strongest interest in Molecular Biology. Took courses in computer science and philosophy that I used in graduate school and in my research. Worked for a summer writing a computer game for predicting the future on the PLATO teaching computer system.
- Graduate student at Caltech, studying the molecular biology of repeated sequences in DNA. Wrote computer programs for measuring protein distributions and DNA reassociation. The first student in the Biology Division to write a thesis solely on a computer.

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Inferring homology – the past 50 years

- 1843 – a definition (similarity in forms and function), but no mechanism
- 1859 – *On the Origin of Species* – a mechanism, *common descent*
- 1960's – 1970's
 - molecular evidence for homology – *structure*: myoglobin/hemoglobin; *sequence*: cytochrome 'c's, trypsin/chymotrypsin
 - statistical approaches (shuffling)
 - scoring methods (minimum evolutionary distance)
- 1970's – 1980's – sequences and sequence databases
 - Needleman-Wunsch, optimal global sequence alignment (1970)
 - mutation data for amino-acid similarity (McLachlan, Dayhoff)
 - private databases and similarity searching (Dayhoff, Doolittle)
 - recombinant DNA (1974)
 - Maxam-Gilbert chemical DNA sequencing (1976)
 - Sanger di-deoxy DNA sequencing (1977)
- 1980's – 2000's – beginning the modern era

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Homology – the beginning

- *Homology* – “the same organ under every variety of form and function”. – Richard Owen, 1843
- Common ancestry is not mentioned, unsurprising for pre-Darwinian and pre-Mendelian times.
- Owen's definition of homology emphasizes structure and location rather than ancestry (structural and functional similarity of a special type).

Fitch, Trends Genet, 2000

What mechanism produces “homology” –
Common Ancestry

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Structures, Sequences, and Homology

- Early inference of "molecular" homology
 - Hemoglobin/Myoglobin Kendrew 1961 (Structure)
 - Cytochromes – Smith & Margoliash, 1964 (sequence)
 - Trypsin/Chymotrypsin Walsh & Neurath, 1964 (sequence)
- Improved methods
 - minimum substitution matrix – Fitch, 1966
 - amino-acid similarity from substitutions – McLachlan, 1971, Dayhoff, 1971

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Winter et al (1968) Science 162:1433

Homology as Applied to Proteins

“Do cats eat bats? Do cats eat bats?” and sometimes “Do bats eat cats?” for you see, as she couldn’t answer either question, it didn’t much matter which way she put it (1).

Our article entitled “Evolution of structure and function of proteases” dealing with the biochemical approach to the subject of evolution as exemplified by studies of proteolytic enzymes (2) put forth a definition of the term “homology” as it applies to similarities in protein structures. This word has been much bandied about and generally used by many to represent a host of ill-defined concepts. We proposed that the word be taken to connote the occurrence of a degree of structural similarity among proteins greater than might be anticipated by chance alone.

significant similarity

This definition has been criticized by Margoliash (3). His position is that since evolution is traditionally the province of the classical biologist, the classical biologist’s definition of “homology” should prevail. This would add to our definition the additional qualification that the protein structures in question must have evolved from a common ancestral gene. The problem with this restrictive definition is that the word, although precisely defined, can seldom be used in a precise sense. For example, did ancestral genes common to divergent populations give rise to “homologous” proteins, or does the occurrence of “homologous” proteins mean that they arose from genes having a common ancestor? It really doesn’t matter how we put it because like Lewis Carroll’s *Alice*, we do not know the answer to either question. The perishable nature of the gene prevents us from obtaining concrete and objective evidence on the nature or existence of ancestral genes.

significant similarity
+ (precise) common ancestry

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Inferring homology – the past 50 years

- 1980’s – 1990 – beginning the modern era
 - *Identification of common molecular subsequences*. Smith and Waterman, 1981. optimal local alignments. (not practical for database searching, > 24 hr for a single search against 3,000 proteins)
 - Genbank – first freely available sequence database (DNA, 1983)
 - Break throughs – oncogenes and viral proteins as kinases and growth factors
 - *Rapid similarity searches of nucleic acid and protein data banks*. Wilbur and Lipman 1983. Heuristic, lookup-based DNA searches.
 - exon shuffling, oncogenes as growth factors
 - *Rapid and sensitive protein similarity searches*. Lipman and Pearson, 1985
 - 1985 – freely available protein databases
 - *Profile analysis* Gribskov et al, (1987)
 - *Improved tools for biological sequence comparison* Pearson and Lipman, 1988 – FASTA
 - *CLUSTAL: a package for performing multiple sequence alignment on a microcomputer*. 1988, Higgins and Sharp
 - *A basic local alignment search tool* Altschul et al., (1990) *BLAST* – statistical thresholds to focus on homologs
 - *Gapped BLAST and PSI-BLAST* Altschul et al. (1997), faster and more sensitive
 - *Complete genomes*

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Perspectives on Homology and Similarity Searching

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- Why (and when) do we infer “Homology” from “similarity” (hint: *excess similarity*)
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Surprises from sequence similarity searches

Proc. Natl. Acad. Sci. USA
Vol. 79, pp. 2836–2839, May 1982
Biochemistry

Viral *src* gene products are related to the catalytic chain of mammalian cAMP-dependent protein kinase

(Rous sarcoma virus/Moloney murine sarcoma virus/transforming protein/protein homologies)

W. C. BARKER AND M. O. DAYHOFF

National Biomedical Research Foundation, Georgetown University Medical Center, Washington D.C. 20007

Proprietary database of ~3,000 protein sequences.
24 hr of VAX780 computer time (\$100/hr)

Simian Sarcoma Virus *onc* Gene, *v-sis*, Is Derived from the Gene (or Genes) Encoding a Platelet-Derived Growth Factor

Doolittle, R.F., Hunkapiller, M.W., Hood, L.E., Devare, S.G.,
Robbins, K.C., Aaronson, S.A. and Antoniades, H.N.
(1983) *Science*, **221**, 275–277.

Proprietary database. (PDP-11??)

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A bit about me - 2

- Post-doctoral fellow at Johns Hopkins (1978), where I went to learn how to clone recombinant DNA. Wrote computer programs for mapping restriction sites and assembling short DNA sequences (match, dmatch)
- Faculty member at U. of Virginia (1983). While waiting for my lab to be set up, worked with David Lipman to write the "FASTP" similarity searching, which became "FASTA", the predecessor to BLAST. Also cloned mouse and human glutathione S-transferases, discovered the human GSTM gene cluster, and the basis for the GSTM1 gene deletion.

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40+ years of rapid sequence comparison

Proc Natl Acad Sci (1983), **80**, 726–730.

Rapid similarity searches of nucleic acid and protein data banks

(global homology/optimal alignment)

W. J. WILBUR AND DAVID J. LIPMAN

Mathematical Research Branch, National Institute of Arthritis, Diabetes, and Digestive and Kidney Diseases, National Institutes of Health, Building 31 Room 4B-54, Bethesda, Maryland 20205

RESEARCH ARTICLE

Science (1985) 227:1435

Rapid and Sensitive Protein Similarity Searches

David J. Lipman and William R. Pearson

FASTP:
2,677 sequences

Proc. Natl. Acad. Sci. USA
Vol. 85, pp. 2444-2448, April 1988
Biochemistry

Improved tools for biological sequence comparison

(amino acid/nucleic acid/data base searches/local similarity)

WILLIAM R. PEARSON* AND DAVID J. LIPMAN†

FASTA:
4,253 sequences

J. Mol. Biol. (1990) 215: 403-410

J. Mol. Biol. (1990) 215:403

Basic Local Alignment Search Tool

Stephen F. Altschul¹, Warren Gish¹, Webb Miller²
Eugene W. Myers³ and David J. Lipman¹

BLAST:
16,524 sequences

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Sequence formats – 1983

Genbank/Genpept:

```

LOCUS      GSTM1_HUMAN          218 aa          linear   PRI 12-SEP-2018
DEFINITION RecName: Full=Glutathione S-transferase Mu 1;
ACCESSION  P09488
...
ORIGIN
    1 mpmilgywdi rglahairll leytdssyee kkytmgdapd ydrsqwlnek fklgldfpnl
    61 pylidgahki tqsnailcyi arkhnlcget eekirvdil enqtdmnhmq lgmicynpfe
    121 eklkpkylee lpeklklyse flgkrpwfag nkitfvdfvl ydvldlhrif epkcldafpn
    181 lkdfisrfeg lekisaymks srflprpvfs kmavwgnk
  
```

EMBL/SwissProt:

```

ID      GSTM1_HUMAN          Reviewed;          218 AA.
AC      P09488; Q5GHG0; Q6FH88; Q8TC98; Q9UC96;
DT      01-JUL-1989, integrated into UniProtKB/Swiss-Prot.
...
SQ      SEQUENCE  218 AA;  25712 MW;  98FB03E87B83A31B CRC64;
        MPMILGYWDI RGLAHAIRLL LEYTDSSYEE KKYTMGDAPD YDRSQWLNEK FKLGLDFPNL
        PYLIDGAHKI TQSNAILCYI ARKHNLGGET EEEKIRVDIL ENQTMDNHMQ LGMICYNPEF
        EKLKPKYLEE LPEKLYSE FLGKRPFAG NKITFVDFLV YDVLDLHRIF EPKCLDAFPN
        LKDFISRFEG LEKISAYMKS SRFLPRPVFS KMAVWGNK
  
```

Dayhoff NBRF/PIR VMS library format:

```

>P1;HAHU
Hemoglobin alpha chain - Human, chimpanzee, and pygmy chimpanzee
VLSPADKTNVKAAWGKVGAGHAGEYGAEALERMFSPPTTKTYFPHFDLSHGSAQVKGHGK
KVADALTNAVAHVDDMPNALSALSDLHAHKLRVDPVNFKLLSHCLLVTLAAHLPAEFTPA
VHASLTKFLASVSTVLTSKYR
  
```

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From NBRF to FASTA format

Dayhoff NBRF/PIR VMS library format:

```

>P1;HAHU
Hemoglobin alpha chain - Human, chimpanzee, and pygmy chimpanzee
VLSPADKTNVKAAWGKVGAGHAGEYGAEALERMFSPPTTKTYFPHFDLSHGSAQVKGHGK
KVADALTNAVAHVDDMPNALSALSDLHAHKLRVDPVNFKLLSHCLLVTLAAHLPAEFTPA
VHASLTKFLASVSTVLTSKYR
  
```



FASTA format:

```

>P1;HAHU Hemoglobin alpha chain - Human, chimpanzee, and pygmy chimpanzee
VLSPADKTNVKAAWGKVGAGHAGEYGAEALERMFSPPTTKTYFPHFDLSHGSAQVKGHGK
KVADALTNAVAHVDDMPNALSALSDLHAHKLRVDPVNFKLLSHCLLVTLAAHLPAEFTPA
VHASLTKFLASVSTVLTSKYR
  
```

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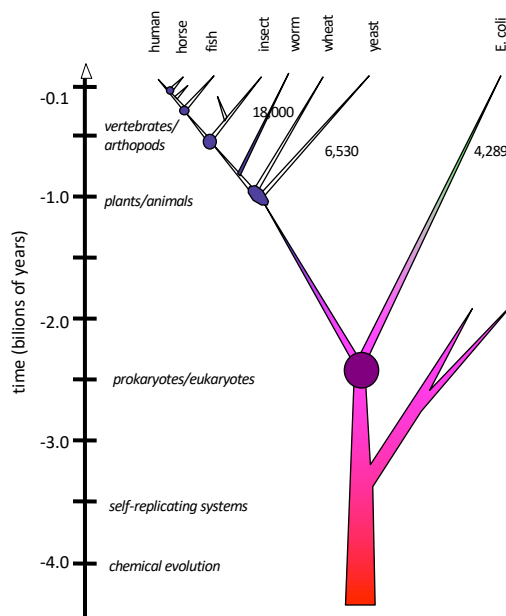
Perspectives on Homology and Similarity Searching

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Homologues share a common ancestor



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Why do sequences look *similar*?
 Why do structures look *similar*?



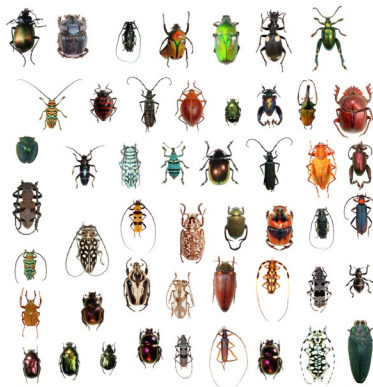
Nebulae look similar because they are shaped by universal physical laws

convergence (not evolution or common ancestry)

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Organisms look similar when they share a common ancestors



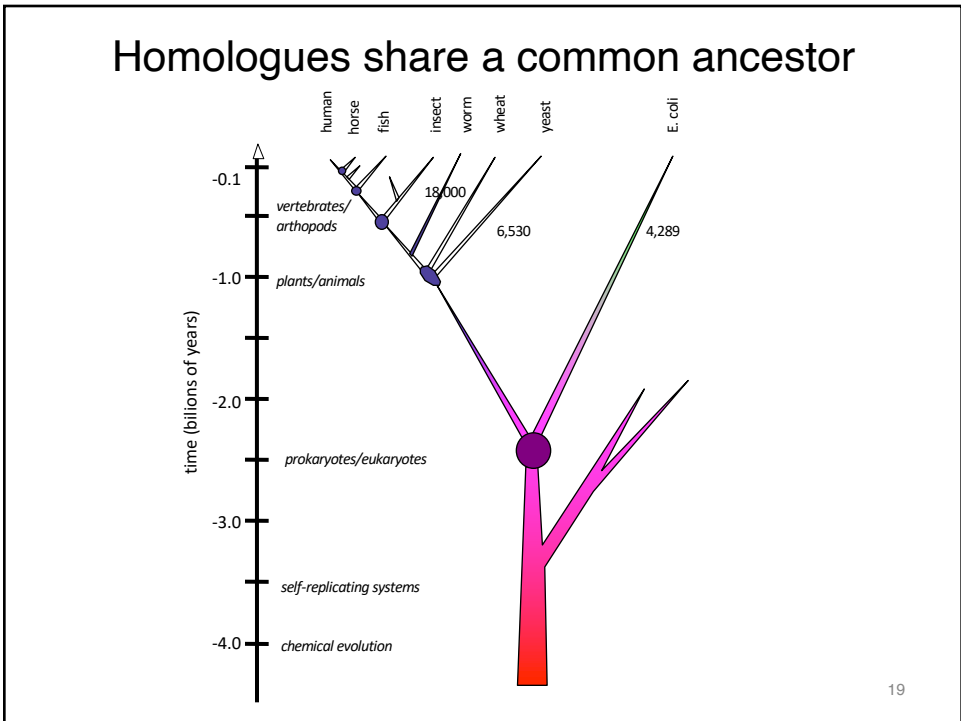
beetle divergence:
 300 Mya



dog diversity:
 15,000 yr

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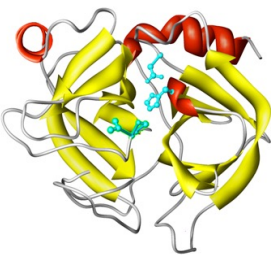
18



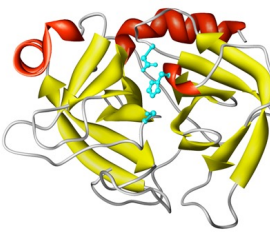
19

Proteins look similar when they share an ancestor

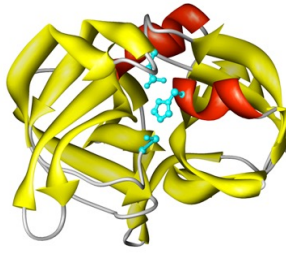
Homology \Leftrightarrow structural similarity
? sequence similarity



Bovine trypsin (5ptp)
Structure: $E() < 10^{-23}$,
RMSD 0.0 Å
Sequence: $E() < 10^{-84}$
100% 223/223



S. griseus trypsin (1sgt)
 $E() < 10^{-14}$ RMSD 1.6 Å
 $E() < 10^{-19}$ 36%; 226/223



S. griseus protease A (2sga)
 $E() < 10^{-4}$; RMSD 2.6 Å
 $E() < 2.6$ 25%; 199/181

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Similarity – homology (divergence) or convergence?

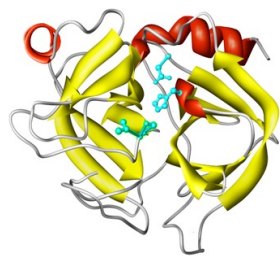
The fundamental assumption of the present approach is that *if the amino-acid sequences of two proteins are so alike that their similarity is very unlikely to have happened by chance, then they will have the same three-dimensional structure and be ancestrally related*. This is based on the finding from X-ray studies that homologous proteins have very similar three-dimensional structures, so that observed amino-acid substitutions usually conserve the folding of the peptide chain. Thus, related proteins remain structurally similar even if the mutation distances are large. ...

One could object to the fundamental assumption, on the grounds that convergent evolution is likely to lead to precisely these kinds of accidental similarities between unrelated proteins. There is not sufficient evidence yet to exclude this possibility. However, no example is yet known where convergent evolution has led to similarities of structure or sequence which approach those found repeatedly in homologous proteins. Rather, the existence of unrelated lysozymes or nucleases, the irregular and apparently random structural features of many proteins, and the large variety of amino-acid substitutions in homologous families of proteins, all suggest that *the number of conceivable ways of evolving an enzyme to perform a given function is astronomically large. Thus, convergent evolution is unlikely to repeat more than a few of the many fine details of structure and sequence in any pair of proteins*.

McLachlan, 1971 *J. Mol. Biol.* 61:409

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When can we infer non-homology?

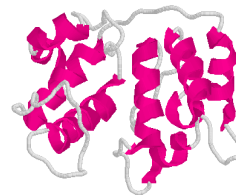


Bovine trypsin (5ptp)
Structure: $E() < 10^{-23}$
RMSD 0.0 Å
Sequence: $E() < 10^{-84}$
100% 223/223

Non-homologous proteins have
different structures



Subtilisin (1sbt)
 $E() > 100$
 $E() < 280$; 25% 159/275

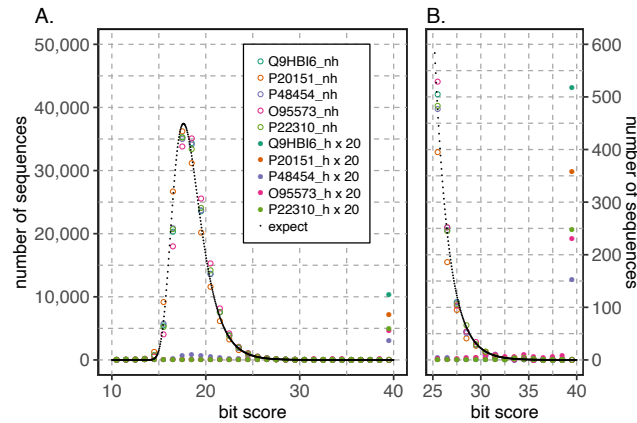


Cytochrome c4 (1etp)
 $E() > 100$
 $E() < 5.5$; 23% 171/190

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Homology inferences are reliable because
similarity statistics are accurate (I)
(we know how unrelated sequences behave)



Distributions of similarity scores in searches with 5 human enzymes. Open circles (_nh) show scores for non-homologs. Closed circles show homolog (_h) scores.

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Inferring Homology from Statistical Significance

- Real **UNRELATED** sequences have similarity scores that are indistinguishable from **RANDOM** sequences
- If a similarity is NOT **RANDOM**, then it must be NOT **UNRELATED**
- Therefore, NOT **RANDOM** (statistically significant) similarity must reflect **RELATED** sequences

With 100+ million protein sequences, it is easy to explore what can happen by chance
Proteins are not statistically similar by chance
Significant similarity implies common ancestry

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Inferring homology – the past 50 years

- 1990's – the genome deluge
 - 1995 *H. influenzae* – first complete bacterial genome
 - *Gapped BLAST and PSI-BLAST* Altschul et al. (1997), faster and more sensitive
 - 1997 – *E. coli*, *S. cerevisiae* genomes
 - 1999 – *C. elegans*
 - 2000 (March) – *D. melanogaster*
 - 2000 (June) – human genome draft sequence

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40+ years of rapid sequence comparison

Proc Natl Acad Sci (1983), **80**, 726–730.

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J. Mol. Biol. (1990) **215**:403

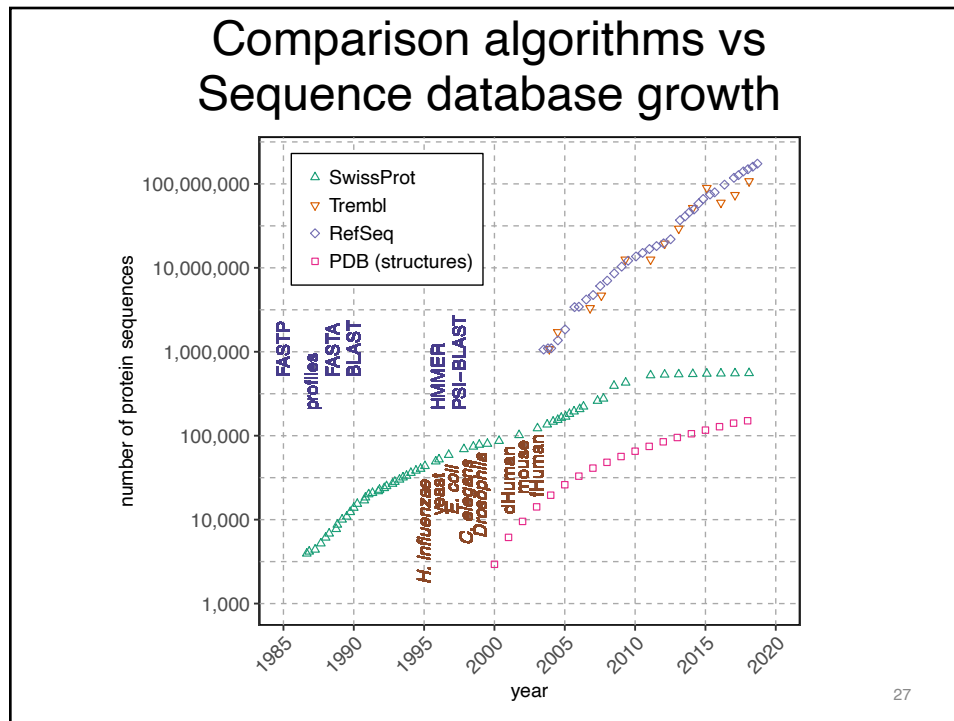
Basic Local Alignment Search Tool

Stephen F. Altschul¹, Warren Gish¹, Webb Miller²
Eugene W. Myers³ and David J. Lipman¹

BLAST:
16,524 sequences

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Inferring homology – the past 50 years

- 1990's – 2000's – BLAST, statistics, and genomes
 - *A basic local alignment search tool* Altschul et al. 1990 fast, sensitive, **built-in statistics**
 - *Maximum-likelihood estimation of the statistical distribution of Smith-Waterman local sequence similarity scores* Mott, 1992
 - 1992 – Yeast chromosome III
 - *Comparison of methods for searching protein sequence databases* Pearson, 1995 (alignments with gaps are better)
 - 1994, 1995 SAM, HMMER Hidden Markov Models for profiles
 - 1995 – H. influenzae genome, 1996 M. jannaschii
 - *Gapped BLAST and PSI-BLAST: a new generation of protein database search programs* Altschul et al, 1997
- 2000 – 202? – the Genome decade(s)
 - Homology and function – orthology, and paralogy

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Inferring Homology – What are the errors?

- False negatives – missing the homologs
 - conservative thresholds (30% identity vs E()-value)
 - searching large databases (reduced sensitivity)
 - using simple models for large families (Pfam families vs clans)
 - distinguishing not-significant from not-homologous

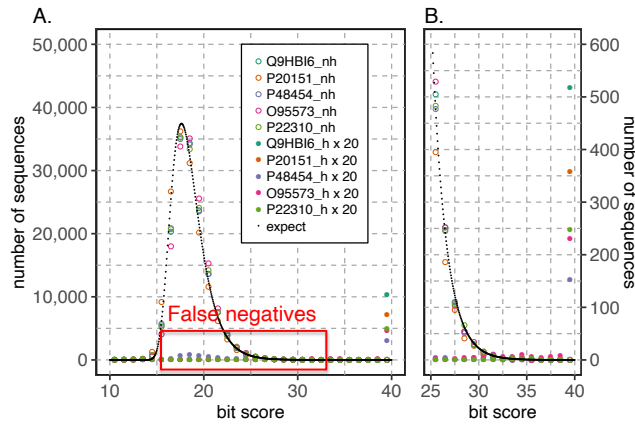
(Only) 14% of environmental (ocean survey) sequences do not share significant similarity with a bacterial reference set

Will *sequence* similarity searching become as sensitive as *structure* comparison?

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Homology inferences are reliable because similarity statistics are accurate (I) (we know how unrelated sequences behave)

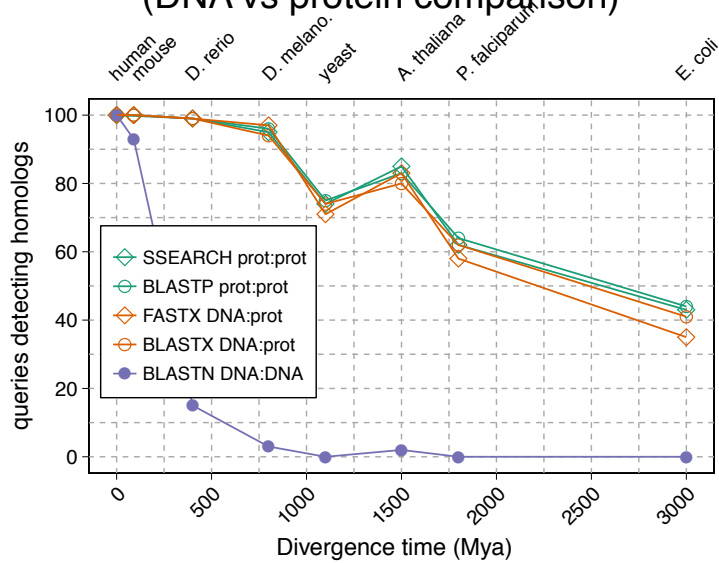


Distributions of similarity scores in searches with 5 human enzymes. Open circles (_nh) show scores for non-homologs. Closed circles show homolog (_h) scores.

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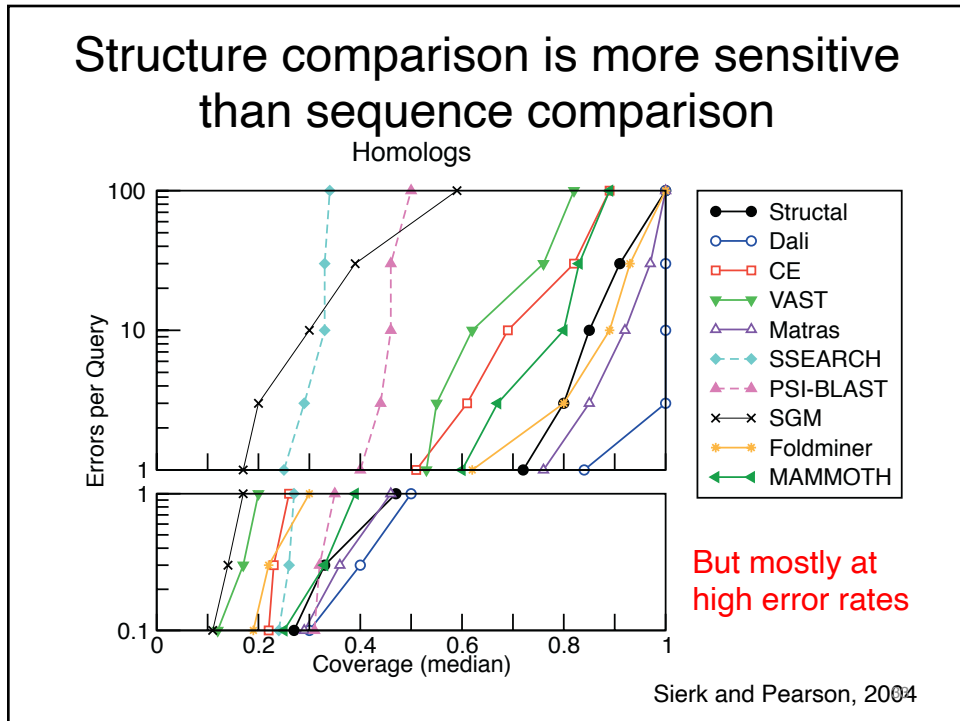
Human enzymes in other organisms (DNA vs protein comparison)



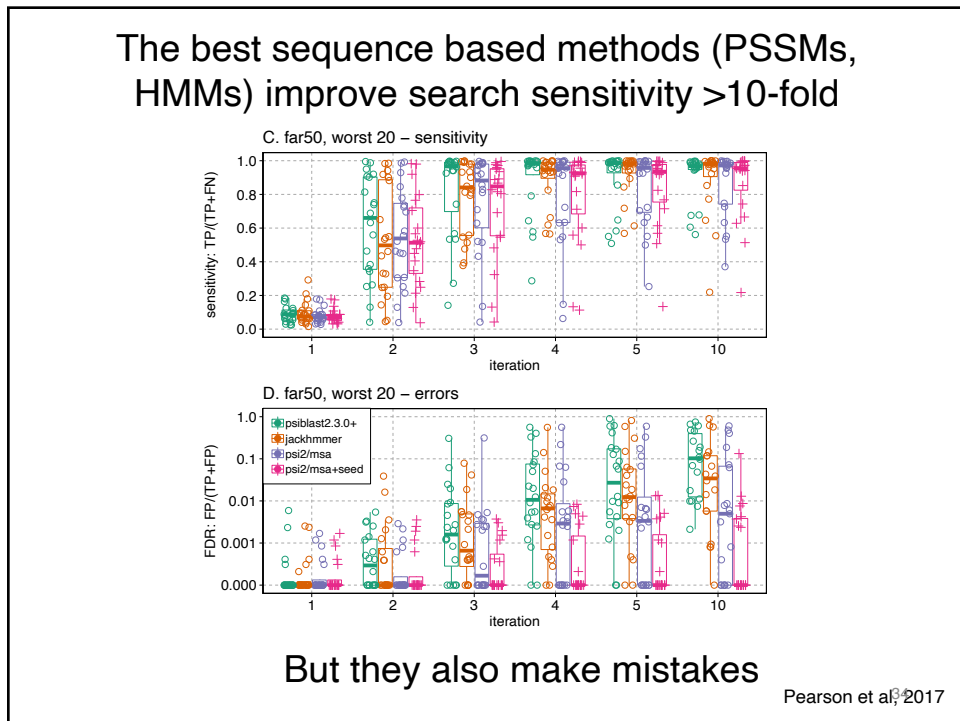
100 protein and mRNA sequences from human enzymes were compared to complete protein and mRNA sets from the indicated organisms.

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Sensitive comparison methods miss homologs

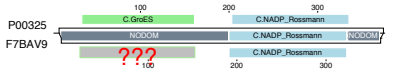
A. ADH1B_HUMAN (P00325) v. ADHX_DROME (P46415)

```
>>ap|P46415|ADHX_DROME Alcohol dehydrogenase class-3 OS=Drosophila melanogaster (379 aa)
qRegion: 34-161; 35-163 : score=379; bits=152.5; Id=0.532; Q=417.8 : C.GroES
qRegion: 203-335;205-339 : score=338; bits=136.0; Id=0.481; Q=368.4 : C.NADP_Rossmann
Region: 2-33 : 3-34 : score=149; bits= 59.9; Id=0.750; Q=141.9 : NODOM
Region: 34-161; 35-163 : score=379; bits=152.5; Id=0.532; Q=417.8 : C.GroES
Region: 162-202;164-204 : score=179; bits= 72.0; Id=0.707; Q=178.6 : NODOM
Region: 203-335;205-339 : score=338; bits=136.0; Id=0.481; Q=368.4 : C.NADP_Rossmann
Region: 349-375;353-379 : score= 96; bits= 38.6; Id=0.519; Q= 78.6 : NODOM
Smith-Waterman score: 1161; 53.3% identity (74.9% similar) in379 aa overlap (2-375:3-379)
```



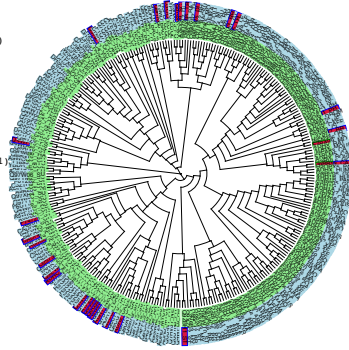
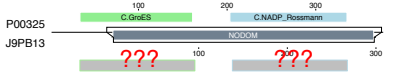
B. ADH1B_HUMAN (P00325) v. F7BAV9_MONDO (F7BAV9)

```
>>|F7BAV9|F7BAV9 MONDO Uncharacterized protein OS=Monodelphis domestica GN (363 aa)
Smith-Waterman score: 1301; 57.5% identity (74.8% similar) in 381 aa overlap (7-373:1-361)
qRegion: 34-161; 28-149 : score=280; bits=113.1; Id=0.537; Q=301.6 : C.GroES
qRegion: 203-335;191-323 : score=545; bits=220.1; Id=0.669; Q=620.5 : C.NADP_Rossmann
Region: 7-202; 1-190 : score=610; bits=246.4; Id=0.619; Q=697.7 : NODOM
Region: 203-336;191-324 : score=548; bits=221.3; Id=0.664; Q=624.3 : C.NADP_Rossmann
Region: 337-373;325-361 : score=143; bits= 57.8; Id=0.568; Q=135.8 : NODOM
```



C. ADH1B_HUMAN (P00325) v. J9PB13_CANLF (J0PB13)

```
>>tr|J9PB13|J9PB13 CANLF Uncharacterized protein OS=Canis lupus familiaris (299 aa)
qRegion: 63-161; 3- 98 : score=243; bits= 96.2; Id=0.554; Q=249.3 : C.GroES
qRegion: 203-335;140-258 : score=242; bits= 95.8; Id=0.445; Q=248.0 : C.NADP_Rossmann
Region: 63-373; 3-297 : score=708; bits=280.4; Id=0.493; Q=793.5 : NODOM
Smith-Waterman score: 708; 45.3% identity (66.8% similar) in 316 aa overlap (63-373:3-297)
```



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Model-based methods miss homologs

A. ADH1B_HUMAN (P00325) v. ADHX_DROME (P46415)

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```

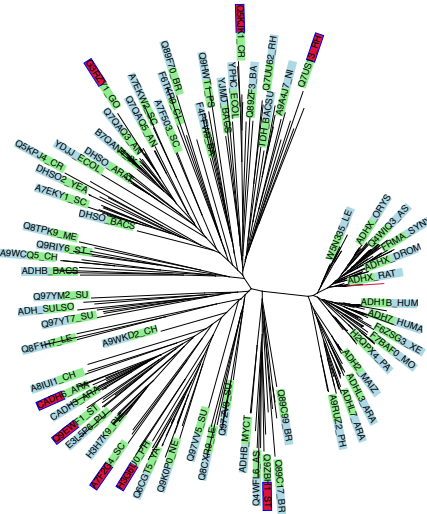
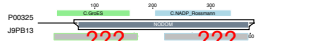


B. ADH1B_HUMAN (P00325) v. F7BAV9_MONDO (F7BAV9)

```
>>|F7BAV9|F7BAV9 MONDO Uncharacterized protein OS=Monodelphis domestica GN (363 aa)
Smith-Waterman score: 1301; 57.5% identity (74.8% similar) in 381 aa overlap (7-373:1-361)
qRegion: 34-161; 28-149 : score=280; bits=113.1; Id=0.537; Q=301.6 : C.GroES
qRegion: 203-335;191-323 : score=545; bits=220.1; Id=0.669; Q=620.5 : C.NADP_Rossmann
Region: 7-202; 1-190 : score=610; bits=246.4; Id=0.619; Q=697.7 : NODOM
Region: 203-336;191-324 : score=548; bits=221.3; Id=0.664; Q=624.3 : C.NADP_Rossmann
Region: 337-373;325-361 : score=143; bits= 57.8; Id=0.568; Q=135.8 : NODOM
```

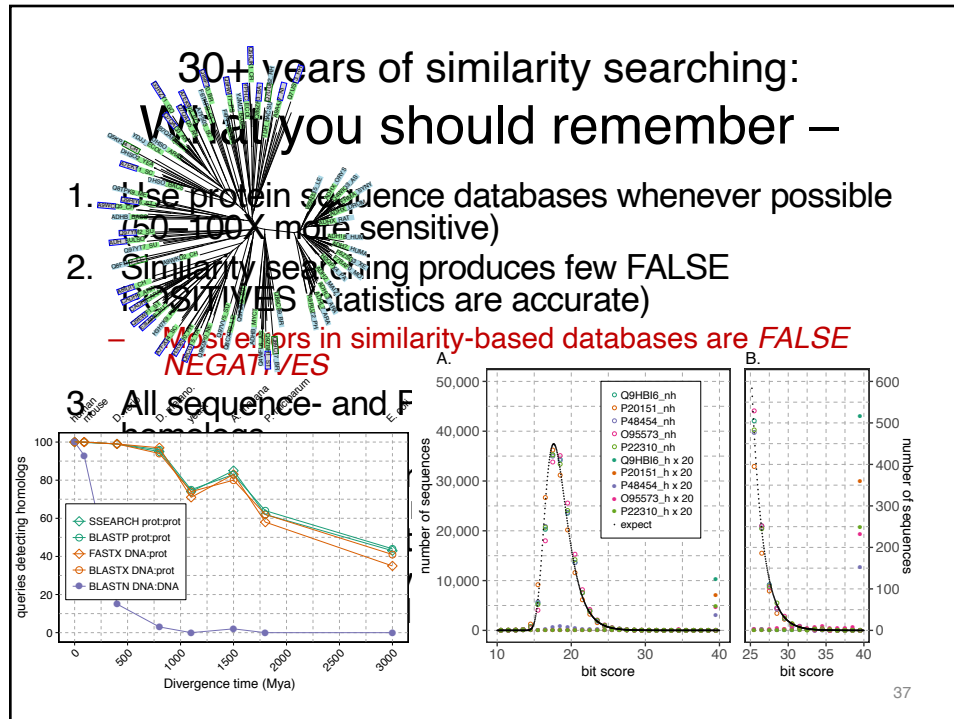


Detection depends on the distance from ~~query~~ query model, the amount of divergence, and the detection threshold

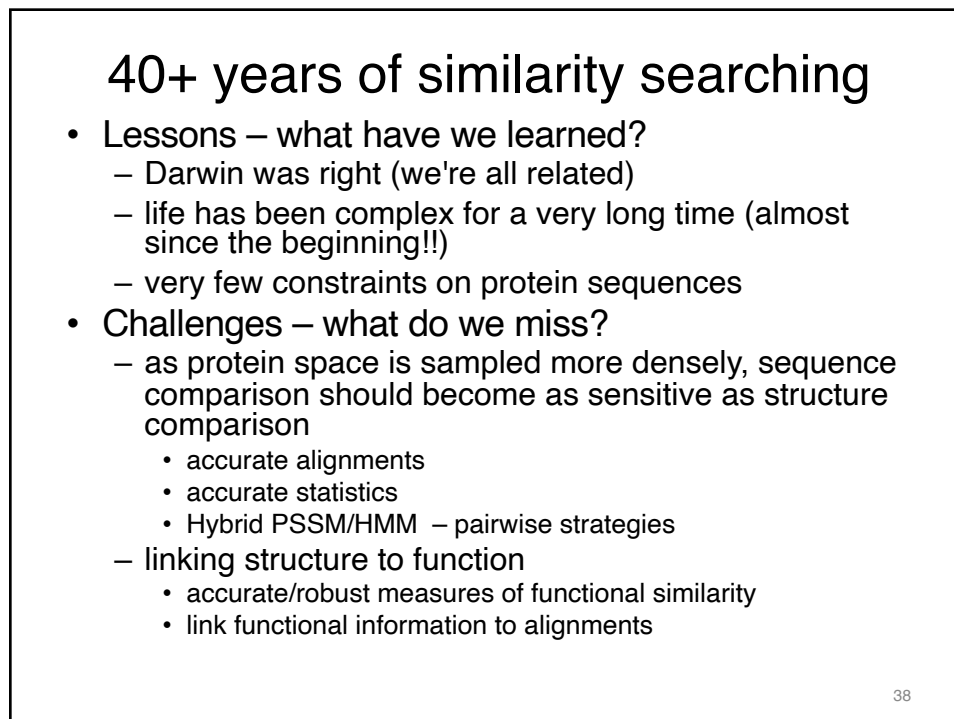


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37



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The challenge of “BIG” data Results should make sense

100% identical sequences behave differently 35 – 60% of the time

But only in different organisms

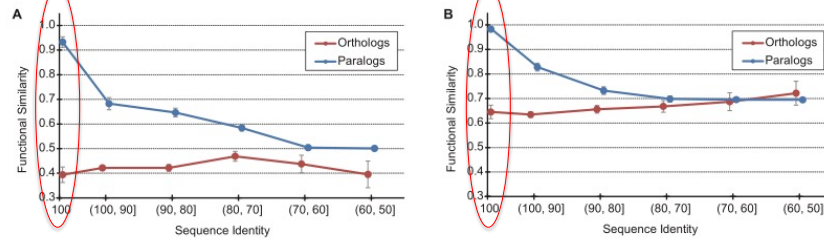


Figure 1. The relationship between functional similarity and sequence identity for human-mouse orthologs (red) and all paralogs (blue). Standard error bars are shown. **(A)** Biological Process ontology, **(B)** Molecular Function ontology. doi:10.1371/journal.pcbi.1002073.g001

PLoS Comput Biol. 2011 7:e1002073.

Testing the ortholog conjecture with comparative functional genomic data from mammals. Nehrt NL, et al.

PLoS Comput Biol. 2012 8:e1002386

On the Use of Gene Ontology Annotations to Assess Functional Similarity among Orthologs and Paralogs: A Short Report. P.D. Thomas et al

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